Aristolane Sesquiterpenes and Highly Brominated Indoles from the Marine Red Alga Laurencia similis (Rhodomelaceae)

by Nai-Yun Ji^a)^b), Xiao-Ming Li^a), Lan-Ping Ding^a), and Bin-Gui Wang^{*a})

a) Institute of Oceanology, Chinese Academy of Sciences, Qingdao 266071, P. R. China (phone: +86-532-82898553; fax: +86-532-82880645; e-mail: wangbg@ms.qdio.ac.cn) b) Graduate School of the Chinese Academy of Sciences, Beijing 100049, P. R. China

Three new polybrominated $1H$ -indoles, compounds $1-3$, and three new aristolane sesquiterpenes,

compounds 4–6, were isolated from the marine red alga *Laurencia similis*, together with seven known natural products. Their structures were elucidated on the basis of detailed spectroscopic and mass-spectrometric analyses, as well as by comparison with literature data.

Introduction. – Marine red algae of the genus Laurencia (Ceramiales, Rhodomelaceae) have been found to be a rich source of structurally unique secondary metabolites. The majority of these metabolites are characterized by the high degree of halogenation, the core structures mainly consisting of sesquiterpenes, diterpenes, triterpenes, and C_{15} acetogenins [1]. The secondary metabolites of the genus vary significantly with species and location. Although the ecological and biological roles of these metabolites have not been clearly elucidated yet, some metabolites have been reported to possess a variety of biological activities such as antifeedant [2], anthelmintic [3], antifouling [4], antimicrobial [4], and cytotoxic [5] properties.

In the course of our ongoing investigations toward biologically active compounds from Chinese marine algae, we examined L. similis collected off the coast of Sanya, Hainan island, China. To the best of our knowledge, only one study has been reported so far concerning the chemical constituents of L . similis [6]. From this species, we isolated and identified five new constituents, including three highly brominated simple $1H$ -indoles, compounds $1-3$, and two new aristolane¹) sesquiterpenes, compounds 4 and 5, as well as the known aristolane 6 [7] [8], isolated herewith for the first time from a natural source. In addition, seven known compounds were obtained from L. similis: 2,3,5-tribromo-1-methyl-1H-indole (7) [9], 2,3,5,6-tetrabromo-1H-indole (8) [6] [9], aristol-1(10)-en-9-one (9) [10], (9 β)-aristol-1(10)-en-9-ol (10) [10], aristol-1(10),8-diene (11) [8] [11], aristol-1,9-diene (12) [7] [8], and elatol (13) [4]. Herein, we report the isolation and structure determination of the new constituents.

Results and Discussion. – The dried and powdered algal material was extracted with a mixture of CHCl₃/MeOH 1:1 (v/v). After solvent removal, the residue was further

¹) Aristolane proper corresponds to (1aR,3aR,7R,7aS,7bS)-decahydro-1,1,7,7a-tetramethyl-1H-cyclopropa[a]naphthalene.

^{© 2007} Verlag Helvetica Chimica Acta AG, Zürich

extracted with 95% aqueous EtOH. The concentrated extracts were combined and partitioned between H_2O and $AcOE$, and the AcOEt-soluble fraction was purified by column chromatography (CC) on silica and *Sephadex LH-20* gel, as well as by preparative thin-layer chromatography (TLC), to yield compounds $1-13$.

Compound 1 was obtained as colorless crystals. Low-resolution EI-MS exhibited a characteristic molecular-ion cluster at m/z 357/355/353/351 in a ratio of 1:3:3:1, which clearly indicated the presence of three Br-atoms. The molecular formula was deduced as $C_8H_4Br_3N$ by positive HR-FAB-MS (*m*/z 353.7954 ([*M* + H]⁺, $C_8H_5{}^{81}Br^{79}Br_2N^+$; calc. 353.7952)). The 13 C-NMR (DEPT) data (*Table 1*) indicated the presence of eight Catoms, including three CH groups and five quaternary C-atoms. The ¹H-NMR spectrum (Table 1) exhibited four signals at $\delta(H)$ 7.22 (d, J=2.6 Hz, H-C(2)), 7.84 (s, H-C(4)), 7.68 (s, $H-C(7)$), and 8.29 (br. s, NH). Detailed comparison with the reported NMR data of the known compounds 7 and 8 [6] [9] suggested that 1 possessed a tribromoindole skeleton. The ${}^{1}H, {}^{1}H$ -COSY spectrum showed a cross-peak between $H-C(2)$ and H-N(1), which indicated that C(2) was connected to N(1). The two *singlets* at $\delta(H)$ 7.84 (H-C(4)) and 7.68 (H-C(7)) confirmed that Br-atoms were attached to C(3), C(5), and C(6), respectively. The HMBC correlations (Fig. 1) from H–C(2) to C(3), C(3a), and C(7a), from H–C(4) to C(3), C(5), C(6), and C(7a), and from H–C(7) to $C(5)$ and $C(3a)$, respectively, confirmed that compound 1 corresponds to 3,5,6-tri b romo-1 H -indole.

Compound 2 was obtained as colorless crystals. Low-resolution EI-MS exhibited a characteristic molecular-ion cluster at m/z 371/369/367/365 in a ratio of 1:3:3:1, indicating three Br-atoms. The molecular formula was deduced as $C_9H_6Br_3N$ by positive HR-FAB-MS (*m*/z 367.8095 ([*M* + H]⁺, C₉H₇⁸¹Br⁷⁹Br₂N⁺; calc. 367.8108)). The ¹³C-NMR (DEPT) spectrum of 2 (*Table 1*) showed the presence of nine C-atoms, with one Me and three CH groups, as well as five quaternary C-atoms. The ¹H-NMR spec-

Position	1		2		3	
	$\delta(H)$	$\delta(C)$	$\delta(H)$	$\delta(C)$	$\delta(H)$	$\delta(C)$
2	7.22 $(d, J=2.6)$	125.2(d)	7.06(s)	129.5 (d)		110.6 (s)
3		91.0(s)		88.7(s)		94.7(s)
3a		127.8(s)	$\overline{}$	128.1(s)		126.6(s)
4	7.84 (s)	123.7(d)	7.81 (s)	123.8 (d)	7.35 $(d, J=8.5)$	120.1(d)
5		116.3 (s)		115.7 (s)	7.29 (dd, $J = 8.5, 1.5$)	124.7(d)
6		118.7 (s)		118.3 (s)		117.1 (s)
	7.68 (s)	116.1 (d)	7.61 (s)	114.4 (d)	7.45 $(d, J=1.5)$	113.6 (d)
7a		134.9 (s)		136.0(s)		136.2 (s)
NH	8.29 (br. s)				8.36 (br. s)	
MeN			3.74(s)	33.3 (q)		

Table 1. ¹H- and ¹³C-NMR Data of 1–3. At 500 and 125 MHz, resp., in CDCl₃. Assignments were corroborated by ¹H,¹H-COSY, HMQC, and HMBC experiments.

trum (*Table 1*) exhibited four *singlets* at $\delta(H)$ 7.06 (H–C(2)), 7.81 (H–C(4)), 7.61 (H– $C(7)$), and 3.74 (Me(10)). Detailed comparison of the spectroscopic data of 1 and 2 suggested that 2 was an N-methylated tribromoindole. Whereas the ${}^{1}H,{}^{1}H$ -COSY spectrum of 2 showed no cross-peaks among the 1 H-NMR signals, the HMBC spectrum (Fig. 1) displayed correlations from H-C(2) to C(3), C(3a), and C(7a), from H-C(4) to C(3), C(5), C(6), and C(7a), from H–C(7) to C(5), C(6), and C(3a), and from H– $C(10)$ to $C(2)$ and $C(7a)$, respectively. These data led to the establishment of the structure of compound 2 as $3,5,6$ -tribromo-1-methyl-1H-indole.

Fig. 1. Selected HMBC correlations for 1–3

Compound 3 was obtained as colorless crystals. Low-resolution EI-MS exhibited a characteristic molecular-ion cluster at m/z 357/355/353/351 in a ratio of 1:3:3:1, which indicated the presence of three Br-atoms. The molecular formula was deduced as $C_8H_4Br_3N$ by positive HR-FAB-MS (*m/z* 353.7934 ([*M*+H]⁺, $C_8H_5{}^{81}Br^{79}Br_2N^+$; calc. 353.7952)), just as for 1. The ¹³C-NMR (DEPT) spectrum of 3 (*Table 1*) showed the presence of eight C-atoms, with three CH groups and five quaternary C-atoms. The ¹H-NMR spectrum of 3 (*Table 1*) exhibited four signals at δ (H) 7.35 (*d*, *J* = 8.5 Hz, $H-C(4)$), 7.45 (d, J=1.5 Hz, H-C(7)), 7.29 (dd, J=8.5, 1.5 Hz, H-C(5)), and 8.36 (br. s, NH). These signals, as well as the corresponding coupling constants, suggested the presence of a benzene ring with an ABX spin system. A comparison with the spectroscopic data of 1 and those of the known compound 7 indicated that 3 was either 2,3,5- or 2,3,6-tribromoindole. A strong HMBC correlation (Fig. 1) between the H-C(4) $(\delta(H)$ 7.35) and C(3) $(\delta(C)$ 94.7) suggested that one Br-atom was bonded to C(6). The other HMBC correlations from H-C(4) to C(6) and C(7a), from H-C(5) to $C(7)$ and $C(3a)$, and from H-C(7) to C(5), C(6) and C(3a) confirmed that the structure of compound 3 corresponds to $2,3,6$ -tribromo-1H-indole.

Compound 4 was obtained as colorless crystals. The IR spectrum indicated the presence of OH (3476) and C=O (1702 $\rm cm^{-1})$ groups. The molecular formula of **4** was determined as $C_{15}H_{24}O_2$, on the basis of positive HR-FAB-MS (m/z 259.1682 ($[M+Na]^+$, $C_{15}H_{24}NaO_2^+$; calc. 259.1673)). However, the ¹³C-NMR (DEPT) spectrum, recorded in CDCl₃, revealed only 14 signals, corresponding to four Me, four CH₂, and three CH groups, as well as three quaternary C-atoms (Table 2). This was due to the fact that the signal of an additional, oxygenated quaternary C-atom was masked by the solvent signals (CDCl₃), as confirmed by recording the spectrum in CD₃OD (*Table 2*), where, indeed, an additional signal was observed at $\delta(C)$ 77.7.

The 1 H-NMR signals of 4 (*Table 2*) could be unequivocally assigned by HMQC experiments, which indicated the presence of an OH group at $\delta(H)$ 2.09 (br. s, 1 H), four Me groups at $\delta(H)$ 1.05 (s), 1.06 (s), 1.20 (s), and 0.94 (d), four CH₂ moieties in the range δ (H) 1.16–3.09, and three CH groups at δ (H) 0.80–1.51. The ¹H₁H₁-H₁ COSY correlations from CH₂(1) to CH₂(2), from CH₂(2) to CH₂(3), from CH₂(3) to $H-C(4)$, and from $H-C(4)$ to Me(14) clearly indicated a $CH_2-CH_2-CH_2-CH$ moiety in 4. The second structural sequence, $CH-CH-CH_2$, was indicated by ${}^{1}H,{}^{1}H-$ COSY correlations from H-C(6) to H-C(7), and from H-C(7) to CH₂(8). The observed key HMBC correlations (Fig. 2) between $H-C(1)$ and both $C(9)$ and C(10), between Me(14) and C(3), C(4), and C(5), between Me(15) and C(4), C(5), $C(6)$, and $C(10)$, and between $H-C(7)$ and $C(9)$ allowed us to connect the above two structural subunits. Furthermore, additional HMBC correlations between Me(12) and C(6), C(7), C(11), and C(13), and between Me(13) and C(6), C(7), $C(11)$, and $C(12)$, respectively, provided the planar structure of 4.

The relative configuration of 4 was determined by a NOESY experiment and by analysis of coupling constants. In the NOESY spectrum, both Me H-atoms, $Me(14)$ and Me(15), showed NOESY correlations with one H-atom of CH₂(1) (δ (H) 1.90 – 1.93), which suggested that the two Me groups were on the same side of the molecular plane. NOESY Correlation between $H - C(6)$ and Me(15) further indicated that $H-C(6)$ had the same orientation as Me(15). The coupling constant between H- $C(6)$ and H-C(7) (J=9.1 Hz) suggested a *cis* orientation [10] [11], as also confirmed by an NOE. The 10-OH group should be *trans* relative to $Me(14)$ and $Me(15)$, since no NOE cross-peak was observed between them. Thus, based on the above evidence, the structure of compound 4 was established as (10β) -10-hydroxyaristolan-9-one.

Fig. 2. Selected HMBC correlations for 4 and 5

Position	4		5			
	$\delta(H)$	$\delta(C)$	$\delta(C)^a$	$\delta(H)$	$\delta(C)$	
$\mathbf{1}$	$1.35 - 1.42$ (<i>m</i>),	29.5 (t)	30.8 (t)		209.9(s)	
	$1.90 - 1.93$ (<i>m</i>)					
2	$1.53 - 1.59$ (<i>m</i>),	22.8 (t)	24.1(t)	$2.29 - 2.42$ (<i>m</i>)	40.5 (t)	
	$1.60 - 1.70$ (<i>m</i>)					
3	$1.16 - 1.25$ (<i>m</i>),	29.1(t)	30.3 (t)	$1.65 - 1.73$ (<i>m</i>),	31.4 (t)	
	$1.31 - 1.35$ (<i>m</i>)			$1.90 - 1.98$ (<i>m</i>)		
4	$1.43 - 1.51$ (<i>m</i>)	34.8 (d)	36.3 (d)	$1.92 - 2.00(m)$	40.1 (d)	
5		46.8 (s)	48.0 (s)		39.1 (s)	
6	$0.80 (d, J=9.1)$	29.0 (d)	31.0 (d)	0.78 $(d, J=8.5)$	35.1 (d)	
7	$1.32 - 1.33$ (<i>m</i>)	22.9(d)	24.8 (d)	1.23 (dd, $J = 8.5, 5.6$)	24.7(d)	
8	2.26 $(d, J=17.0)$,	34.5 (t)	35.7(t)	$6.00 - 6.04$ (<i>m</i>)	127.3(d)	
	3.09 (dd, $J=17.0, 9.0$)					
9		215.9(s)	219.7(s)	5.86 (dd, $J=10.1, 1.9$)	118.8 (d)	
10		$n.d.^{b}$	77.7 (s)	2.58 (br. s)	54.7 (d)	
11		21.0(s)	21.6(s)		26.1(s)	
12	1.05(s)	16.8 (q)	17.3 (q)	1.01(s)	16.0 (q)	
13	1.06(s)	30.6 (q)	31.0 (q)	1.16(s)	30.5 (q)	
14	0.94 $(d, J=6.7)$	17.0 (q)	17.5 (q)	1.05 $(d, J=6.6)$	15.5 (q)	
15	1.20(s)	18.4 (q)	18.8 (q)	0.56(s)	15.9 (q)	
OH	2.09 (br. s)					
	\mathbb{R} . Le CD OD \mathbb{R} . Not detected due to express consider					

Table 2. ¹H- and ¹³C-NMR Data of 4 and 5. At 500 and 125 MHz, resp., in CDCl₃ (unless noted otherwise). Assignments were corroborated by ${}^{1}H, {}^{1}H$ -COSY, HMQC, and HMBC experiments.

 α) In CD₃OD. α) Not detected due to solvent overlap.

Compound 5 was obtained as a colorless oil. Its IR spectrum indicated the presence of C=O (1720) and C=C (1631 cm⁻¹) functions, and the molecular formula was determined as C₁₅H₂₂O by positive HR-FAB-MS (m/z 241.1564 ($[M+Na]^+$, C₁₅H₂₂NaO⁺; calc. 241.1568)). The ¹³C-NMR (DEPT) spectrum of 5 (*Table 2*) showed the presence of 15 C-atoms, corresponding to four Me, two $CH₂$, and six CH groups, as well as three quaternary C-atoms. The ${}^{1}H, {}^{1}H$ -COSY correlations from CH₂(2) to CH₂(3), from CH₂(3) to H–C(4), and from H–C(4) to Me(14) indicated a spin systems comprising a $CH_2-CH_2-CH-Me$ unit. A second system was derived from correlations from H–C(6) to H–C(7), from H–C(7) to H–C(8), from H–C(8) to H–C(9), and from $H-C(9)$ to $H-C(10)$, in accord with the presence of a $CH-CH-CH-CH$ moiety. The HMBC correlations (Fig. 2) between $CH₂(2)$ and C(1) and C(10), between $H-C(9)$ and $C(1)$, between Me(14) and $C(3)$, $C(4)$, and $C(5)$, and between Me(15) and $C(4)$, $C(5)$, $C(6)$, and $C(10)$, respectively, established the connectivity of the above two sequences at $C(1)$ and $C(5)$. Additional HMBC correlations between Me(12) and $C(6)$, $C(7)$, $C(11)$, and $C(13)$, as well as between Me(13) and $C(6)$, $C(7)$, $C(11)$, and $C(12)$, allowed the establishment of the planar structure of 5.

Based on biogenetic considerations, in combination with analysis of coupling constants and COSY experiments, the configurations at $C(4)$, $C(5)$, $C(6)$, and $C(7)$ of 5 were established to be identical with those in 4. The coupling constant between H- $C(6)$ and H-C(7) (J=8.5 Hz) agreed with a *cis* orientation of H-C(6) and H-C(7)

[10] [11]. The ¹H,¹H-COSY cross-peak between $H - C(10)$ and Me(15) indicated a longrange W-type coupling, which suggested that $H-C(10)$ and Me(15) were trans-oriented [12]. Notably, compound 5 differs from the known compound aristol-9-en-1-one only in the position of the C=C bond [7]. Thus, from the above data, the structure of 5 was determined as aristol-8-en-1-one.

Compound 6, a colorless oil, was identified as aristol-9-en-1-one. Interestingly, this compound had been previously obtained synthetically by oxygenation of (1α) -aristol-9en-1-ol with $CrO₃$, or by photooxygenation of calarene [7] [8]. Our report shows for the first time that 6 is also a natural product.

Previous investigation toward the metabolites of the marine red alga species L. similis was less than other species, such as L . *obtusa*, L . *majuscula*, L . *nipponica*, and L . okamurai, within the genus Laurencia. To the best of our knowledge, only two species from this genus, *i.e.*, L . *similis* and L . *brongniartii*, have been reported to contain polybrominated indole metabolites. These may be produced via similar biogenetical pathways, indicating a close affinity between L. similis and L. brongniartii. Our results are unusual in the sense that both halogenated indole alkaloids and sesquiterpenes are cooccurring in L. similis, aristolane-type sesquiterpenes without halogen substitution being peculiar in the metabolites of the genus *Laurencia*.

This work was financially supported by the National Natural Science Foundation of China (No. 30530080) and by the Department of Science & Technology of Shandong Province (2006GG2205023). The program of Bairen Jihua from the Chinese Academy of Sciences (awarded to B.-G. W.) is gratefully acknowledged.

Experimental Part

General. Column chromatography (CC): silica gel (200-300 and 300-400 mesh; *Qingdao Haiyang* Chemical Group, Co.) or Sephadex LH-20 (Sigma). TLC: precoated silica-gel plates GF-254 (Qingdao Haiyang). Optical rotation: Atago Polax-L polarimeter. UV/VIS Spectra: Beckman DU-640 spectrophotometer; λ_{\max} (log ε) in nm. IR Spectra: *Nicolet Nexus-470* spectrophotometer; in cm⁻¹. ¹H- and ¹³C-NMR Spectra: Bruker Avance-500 spectrometer; at 500/125 MHz, resp.; δ in ppm, J in Hz. Low- and high-resolution FAB-MS: VG Autospec-3000 mass spectrometer; in m/z.

Algal Material. The red alga L. similis was collected off the coast of Hainan island, P. R. China, in March 2006, and identified by Prof. Lan-Ping Ding, Institute of Oceanology, Chinese Academy of Sciences (IOCAS). A voucher specimen (06M04) was deposited at the Key Laboratory of Experimental Marine Biology of IOCAS.

Extraction and Isolation. The dried and powdered alga L. similis (487 g) was extracted with CHCl₃/ MeOH 1:1. After solvent removal, the residue was further extracted with 95% aq. EtOH. The concentrated extracts were combined and partitioned between H2O and AcOEt. The AcOEt-soluble fraction was purified by CC (SiO₂; petroleum ether (PE)/AcOEt gradient) to afford nine fractions: Fr. 1–Fr. IX. Fr. I (eluted with PE) was further purified by prep. TLC (cyclohexane) to afford 7 (56.8 mg), and a mixture of 11 and 12 (31.7 mg). Fr. II (eluted with PE/AcOEt 100:1) was purified by prep. TLC (PE/AcOEt 100:1) to afford 2 (8.0 mg). Fr. III (eluted with PE/AcOEt 50:1) was further separated by CC (SiO₂ and Sephadex LH-20; CHCl₃/MeOH 1:1) followed by prep. TLC to afford 1 (8.8 mg), 3 (15.0 mg), 5 (29.8 mg), 6 (5.0 mg), 8 (56.0 mg), 9 (3.0 mg), 10 (103.8 mg), and 13 (8.7 mg). Fr. VI (eluted with PE/AcOEt 10:1) was further separated by CC (SiO₂ and Sephadex LH-20; CHCl₃/MeOH 1:1) followed by prep. TLC to afford 4 (6.5 mg).

3,5,6-Tribromo-1H-indole (1). Colorless crystals. M.p. 123-124°. UV (CHCl₃): 218 (5.15), 241 (4.64). IR (KBr): 3452, 1641, 1613, 1447, 1411, 1262, 1094, 1019, 799. ¹H- and ¹³C-NMR: see *Table 1*. EI-MS: 357 (31)/355 (94)/353 (100)/351 (38) (M⁺); 276 (29)/274 (58)/272 (33); 195 (44)/193 (47); 114 (33). HR-FAB-MS (pos.): 353.7954 ($[M+H]^+$, $C_8H_5^{81}Br^{79}Br_2N^+$; calc. 353.7952).

3,5,6-Tribromo-1-methyl-1H-indole (2). Colorless crystals. M.p. 166-168°. UV (CHCl₃): 218 (5.17), 240 (4.51). IR (KBr): 1649, 1607, 1512, 1462, 1417, 1259, 1122, 1082, 802. ¹H- and ¹³C-NMR: see *Table* 1. EI-MS: 371 (31)/369 (95)/367 (100)/365 (37) (M^+) ; 356 (2)/354 (6)/352(6)/350 (2) $([M-Me]^+)$; 290 $(10)/288$ $(21)/286$ (11) ; 209 $(41)/207$ (44) ; 128 (46) . HR-FAB-MS (pos.): 367.8095 $([M+H]^+,$ $C_9H_7^{81}Br^{79}Br_2N^+$; calc. 367.8108).

2,3,6-Tribromo-1H-indole (3). Colorless crystals. M.p. 105-107°. UV (CHCl₃): 216 (5.15), 240 (4.44). IR (KBr): 3380, 1701, 1611, 1503, 1435, 1220, 1048, 809. ¹H- and ¹³C-NMR: see *Table 1*. EI-MS: 357 (30)/ 355 (92)/353 (100)/351 (34) (M⁺); 276 (23)/274 (47)/272 (25); 195 (17)/193 (17); 114 (34). HR-FAB-MS (pos.): 353.7934 ($[M + H]^+$, C₈H₅⁸¹Br⁷⁹Br₂N⁺; calc. 353.7952).

(10 β)-10-Hydroxyaristolan-9-one (4). Colorless crystals. M.p. 94-96°. $[\alpha]_D^{26} = +11.4$ (c=0.25, CHCl₃). UV (CHCl₃): 210 (4.98), 239 (4.14). IR (KBr): 3476, 2959, 2936, 1702, 1091, 1014. ¹H- and ¹³C-NMR: see *Table 2*. FAB-MS (pos.): 236 (M^{+}), 219 ($[M-H_2O+1]^+$). HR-FAB-MS (pos.): 259.1682 $([M+Na]^+, C_{15}H_{24}NaO_2^+;$ calc. 259.1673).

Aristol-8-en-1-one (5). Colorless oil. UV (CHCl₃): 212 (4.94), 240 (4.13). $[a]_D^{26} = +46.0$ ($c = 1.07$, CHCl₃). IR (KBr): 2962, 2873, 1720, 1631, 1263, 1081, 1018. ¹H- and ¹³C-NMR: see *Table 2*. FAB-MS (pos.): 218 (M^+) , 175. HR-FAB-MS (pos.): 241.1564 $([M+Na]^+, C_{15}H_{22}NaO^+$; calc. 241.1568).

REFERENCES

- [1] K. L. Erickson, in 'Marine Natural Products', Ed. P. J. Scheuer, Academic Press, New York, 1983, Vol. 5, p. 131.
- [2] K. Kurata, K. Taniguchi, Y. Agatsuma, M. Suzuki, Phytochemistry 1998, 47, 363.
- [3] D. Davyt, R. Fernandez, L. Suescun, A. W. Mombrú, J. Saldaña, L. Damínguez, J. Coll, M. T. Fujii, E. Manta, J. Nat. Prod. 2001, 64, 1552; G. Topcu, Z. Aydogmus, S. Imre, A. C. Gören, J. M. Pezzuto, J. A. Clement, D. G. I. Kingston, J. Nat. Prod. 2003, 66, 1505.
- [4] G. M. König, A. D. Wright, J. Nat. Prod. 1997, 60, 967.
- [5] E. G. Juagdan, R. Kalidindi, P. Scheuer, Tetrahedron 1997, 53, 521; J. Sun, D. Shi, M. Ma, S. Li, S. Wang, L. Han, Y. Yang, X. Fan, J. Shi, L. He, J. Nat. Prod. 2005, 68, 915.
- [6] M. Masuda, S. Kawaguchi, Y. Takahashi, K. Okamoto, M. Suzuki, Bot. Mar. 1999, 42, 199.
- [7] G. Rücker, U. Kretzuschmar, Liebigs Ann. Chem. 1971, 748, 214.
- [8] H. Takeshita, I. Shimooda, T. Hatsui, *Bull. Chem. Soc. Jpn.* **1980**, 53, 3721.
- [9] G. T. Carter, K. L. Rinehart Jr., L. H. Li, S. L. Kuentzel, J. L. Connor, Tetrahedron Lett. 1978, 19, 4479.
- [10] L. Shide, A. Olbrich, R. Mayer, G. Rücker, Planta Med. 1987, 53, 556.
- [11] G. Vidari, Z. Che, L. Garlaschelli, Tetrahedron Lett. 1998, 39, 6073.
- [12] K. L. Erickson, J. A. Beutler, G. N. Gray, J. H. Cardellina II, M. R. Boyd, J. Nat. Prod. 1995, 58, 1848.

Received November 24, 2006